REGENERATIVE TREATMENT OF AN IMMATURE TOOTH WITH APICAL PERIODONTITIS USING PLATELET-RICH FIBRIN AND MM-MTA: A CASE REPORT

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Abstract

This case report describes the treatment of a necrotic immature permanent central incisor with symptomatic apical periodontitis, which was not treated with conventional apexification techniques. Instead, a regenerative approach based on the literature’s methods for regeneration was provided. The root canal was gently debrided of necrotic tissue with a large diameter file, irrigated with 5,25% NaOCl and then medicated with 2% chlorhexidine. After 21 days, the tooth was asymptomatic. The canal was accessed and irrigated with saline solution and a tissue scaffold was created with PRF (Platelet-Rich Fibrin) prepared using centrifugation. The final restoration was completed immediately with MM-MTA and a composite resin. The tooth was controlled every three months over a period of 2 years and has remained symptom free. Radiographic examination shows healing of the apical radiolucency suggesting that appropriate biologic responses can occur with this type of treatment of the necrotic immature permanent tooth.

Keywords: Platelet-rich fibrin - regeneration - immature tooth - apical periodontitis.

Résumé

Ce rapport de cas décrit le traitement d'une incisive centrale permanente immature atteinte de nécrose pulpaire compliquée d'une parodontite apicale symptomatique non traitée par les techniques classiques d’apexification. Une approche régénératrice basée sur les méthodes de régénération décrites dans la littérature a été appliquée. Le canal radiculaire a été débridé du tissu nécrotique avec une lime de grand diamètre, irrigué avec 5,25% de NaOCl et ensuite rempli d'un produit antiseptique à base de chlorhexidine 2%. Vingt et un jours plus tard la dent était complètement asymptomatique. L'accès au canal a donc été effectué ainsi qu’une irrigation avec une solution saline et la pose d'une matrice tissulaire de et la création d'une matrice tissulaire de FRP (fibrine riche en plaquettes) préparée par centrifugation. La restauration finale a été complétée immédiatement avec MM-MTA et une résine composite. La dent a été contrôlée tous les trois mois sur une période de 2 ans et est restée asymptomatique. L'examen radiographique a montré la guérison de la radiolucité apicale suggérant que des réponses biologiques appropriées pouvaient se produire avec ce type de traitement au niveau d’une dent permanente immature nécrosée.

Mots clés : fibrine enrichie en plaquettes - régénération - dent immature - parodontite apicale.
Introduction

Trauma and/or carious exposure in immature permanent teeth can lead to pulp necrosis and arrested root development [11]. The consequences of interrupted development include a poor crown/root ratio, very thin root canal walls, an increased risk of root fracture and an open apex [2]. It is reported in the literature that 25% of schoolchildren suffer dental trauma while 33% of adults have experienced trauma affecting the permanent dentition with a majority of accidents occurring before 19-years-old [3]. It is important to emphasize that these cases are often poorly managed and the pulp necrosis promotes the development of apical periodontitis. Traditionally, the open apex is managed by a root end closure technique using calcium hydroxide (apexification) [4, 5]. More recently, the placement of mineral trioxide aggregate as an apical barrier has been advocated [6, 7]. Whilst both of these techniques have been shown to have good long-term success [8, 9] in the absence of continued root development, they do not help strengthen the root, rendering it prone to fracture.

The ideal treatment of these immature teeth with necrotic pulps and apical periodontitis (symptomatic or not) would be to stimulate the regeneration of the pulp-dentin complex to obtain further root development [10, 11]. Regenerative endodontic procedures are defined as biologically based procedures designed to replace damaged cells and structures with live viable tissues, which restore the normal physiologic functions [12]. Regeneration can be achieved through the activity of cells from the pulp, periodontium, vascular or immune systems [12] or alternatively through stem cells therapies [13]. There are numerous case reports and series in the literature reporting the use of regeneration/vasculization procedures to treat immature permanent teeth with necrotic pulps with or without apical periodontitis [1, 2, 14 - 21]. These case reports and series have demonstrated radiographical evidence of increased root length and thickening of the root walls with apical closure.

During the year 2001, Choukroun introduced the Platelet-Rich Fibrin (PRF) to be used for guided bone regeneration procedures around implants and sinus lifts [22 - 24]. This seemed interesting in surgery and attempts were made to regenerate the pulp of immature teeth using the PRF as a scaffold to provide a spatially correct position of stem cells with growth factors [16].

In this case report we describe and discuss the use of PRF covered with MM-MTA (Micro-Mega, Besançon, France) to stimulate the continuous root development of a necrotic immature tooth with symptomatic apical periodontitis.

Case report

A 10-year-old girl was referred complaining of pain from the maxillary left central incisor [21]. The pain had been present for several days. There was a history of trauma some 3-4 months previously but the patient did not seek any treatment at the time. There was no relevant medical history. Clinical examination revealed redness of the gingival tissues. The tooth was tender to percussion. Thermal test was negative. Specifically, there was no response to Dry ice (CO2 ICE / Odontest Pulp Thermal test, Miltex New York, NY, USA). Once the working length was measured using the same 80 K-file (Dentsply-Maillefer) and a Root ZX mini apex locator (J Morita, MFG Corp., Kyoto, Japan) revealed periapical disease with an incomplete root development and a very wide-open apex (Fig. 1). The tooth was diagnosed with a symptomatic apical periodontitis associated with a necrotic pulp. The treatment plan was to explore the possibility of carrying out a regeneration procedure in two visits using PRF and MM-MTA (Micro-Mega MTA, Besançon, France) after root canal disinfection with 5, 25% sodium hypochlorite and 2% chlorhexidine gluconate. The treatment plan was agreed and appropriate consent obtained.

Local anesthetic infiltration was given using 3% Scandicaine without epinephrine (Mepivacaine Septodont, Saint-Maur des Fosses, France) after the alveolar mucosa was smeared with a topical anesthetic gel (20% Benzocaine, Sultan) over tooth #21. Under rubber dam isolation (Hygienic Coltene/Whaledent, Germany) the pulp chamber was accessed using a round diamond bur and an Endo-Access bur (Dentsply Maillefer, Ballaigues, Switzerland) in an air turbine hand-piece with copious irrigation (NSK, PAN Air, Nakaniishi Inc., Tochigi-Ken, Japan).

The canal was gently debrided using one manual shaping file (80 K-file, Dentsply Maillefer, Ballaigues, Switzerland) and irrigated with a solution of sodium hypochlorite 5.25% to flush out gross debris using a 3-mL Leuer Lock syringe with a 27-G irrigating needle tip (Monoject endodontic irrigating syringe Covidien, Henry Schein dental, USA). The canal length was measured using the same 80 K-file (Dentsply-Maillefer) and a Root ZX mini apex locator (1 Morita, MPG Corp., Kyoto, Japan). Once the working length was established and confirmed by a radiograph, the canal was irrigated with 18 ml of NaOCl (Clorox 5.25%, Spartan, Egypt) and dried with sterile absorbent paper points (Coarse, Dentsply Maillefer). It was then filled with 2% Chlorhexidine (Calaplast) for three weeks. A sterile cotton pellet was covered with 4 mm of Cavit G (Espe, Seefeld, Germany) to provisionally seal the access cavity.

A second appointment was set 3 weeks later. During this session, preparation of PRF was performed. Venous blood (10 mL) was transferred to the tubes, which were centrifuged at 2700 rpm for 12 minutes. Initial centrifugation produced 3 distinct layers, namely a layer of red blood corpuscles and leukocytes at the bottom, a layer of platelet concentrate, fibrin clot and a top layer, the supernatant, which is the cell-free plasma, acellular plasma or platelet-poor plasma. The absence of anticoagulant allows massive and
Fig. 1: Pre-operative x-ray (February 2014).

Fig. 2a: PRF in the tubes after centrifugation.

Fig. 2b: PRF extracted with the tweezers.

Fig. 3: X-ray taken after second session and setting of the PRF and MM-MTA.

Fig. 4: X-ray taken after 3 months.

Fig. 5: X-ray taken after 6 months.

Fig. 6: After 18 months.
rapid platelet activation and initiation of coagulation. Once separated from the red cells, the clot has to be removed and the operation must be fast, otherwise, coagulation begins resulting in an unstable polymerization of fibrin (Fig. 2).

After irrigating the canal with 9 mL of sterile saline to remove the Chlorhexidine, the canal was dried with sterile paper points and the PRF was introduced in the canal. A layer of MM-MTA (4 mm) covered the PRF and a wet cotton pellet placed over for 15 minutes to allow the initial setting of the material. The tooth was then restored with a composite (3M ESPE Filtek Z250 XT, 2510 Conway Avenue St. Paul, MN 55144-1000 USA) during the same session.

Radiographic controls were taken on a 3-months basis (3 months / 6 months, until 2 years), to assess apical closure and continuous root edification, decrease of the periapical radiolucency, and thickening of the root canal walls.

Discussion

Tissue regeneration using revascularization to induce apexogenesis rather than tissue replacement using artificial substitutes is a relatively new treatment modality for infected, immature teeth with necrotic pulps [16].

According to some authors [19, 25, 26], there are three key steps to revascularization /regeneration procedures: (i) disinfecting the canal system, (ii) inducing bleeding to form a blood clot or placement of a suitable matrix into which new tissue can grow and (iii) a bacteria-resistant coronal seal [1].

A necrotic immature tooth has an infected canal space with very thin and divergent canal walls. Mechanical instrumentation is not indicated in these cases because of this fragile and delicate structure, which might lead to fracture and jeopardize the future of the tooth [27].

However, controlling infection in the canal space is an important initial step in the revascularization/regeneration process [1]. It is also suggested that a higher level of disinfection is required for pulp regeneration. Two in vitro studies conducted by Hoshino et al. and Sato et al. [28, 29] showed that the mixture of ciprofloxacin, metronidazole and minocycline was effective against endodontic pathogens and was able to disinfect into the dentine tubules. This antibiotic mixture has become known as Hoshino’s paste.

Both Windley et al. and Cohenca et al. [30, 31] have shown that irrigation with sodium hypochlorite followed by triple antibiotic paste renders 70-78% of the canals culture negative. This protocol has become the basis for disinfection in revascularization cases. Most of the studies used Hoshino’s triantibiotic paste as the intracanal medicament of choice in the disinfection process for the treatment of nonvital immature teeth undergoing revascularization treatment [2, 10, 17, 21, 25, 32]. However, there are issues regarding possible sensitivity and bacterial resistance to some of the used medicaments [31, 33], and staining or possible discoloration of the tooth has been reported [19], so others have used calcium hydroxide [25] and one study used formocresol [16].

Calcium hydroxide, which has been extensively used in apexification procedures [34], has antibacterial properties due among others to its high pH, but the proteolytic action of calcium hydroxide tends to reduce the organic support due to disruption of links between collagen fibers and hydroxyapatite crystals. This reduction in microhardness of dentine may impact adversely on dentine fracture resistance [5, 35].

2% chlorhexidine (CHX) was used as a temporary medication in the present case. Chlorhexidine is an antibacterial...
agent with a broad spectrum, effective against both E. faecalis and Candida albicans like 5.25% sodium hypochlorite solution [36]. Its antibacterial properties are due to an alkaline pH and a continuous release of OH- ions [37]. In addition, chlorhexidine is adsorbed and released by dental tissues, this long-term antibacterial activity is known as ‘substantivity’ and has been demonstrated in vitro when chlorhexidine has been used in association with different vehicles i.e. liquids, gels, or devices for controlled release [38].

One of the mechanisms that may explain its effectiveness is based on the interaction between the positive charge of the molecule and the negative charged phosphate groups present on the bacterial cell wall, which allows CHX molecule to penetrate inside the bacteria [39, 40].

Paquette (2007) [41] showed that 2% chlorhexidine gluconate solution used as intracanal medication for 7 to 15 days reduced bacterial concentration and Gomes (2009) [42] added that CHX was able to diffuse into the dentine tubules and reach the outer surface of the root, exerting its antimicrobial activity.

Subsequent to disinfection, an appropriate scaffold is needed to promote cell growth and differentiation [15]. Thibodeau et al. [14] showed that roots containing a blood clot after disinfection had better treatment outcomes in a dog model than those that did not have a blood clot in the apical part of the canal. It is considered that the presence of blood clot in canal space provides a fibrin scaffold that would promote the regeneration of tissue in root canal space (10, [14]). However, the induction and maintenance of a blood clot is extremely difficult in revascularization procedures and appears to be its limitation [17].

Platelet-rich fibrin, a second-generation platelet concentrate can be considered as an autologous healing biomaterial incorporating leucocytes, platelets and wide range of key healing proteins in a dense fibrin matrix [43]. Unlike platelet-rich plasma, PRF preparation eliminates the redundant process of adding anticoagulants or bovine thrombin making it similar to centrifuged natural blood. Furthermore, a highly resistant and elastic membrane of fibrin is obtained [44], which allows a slow continuous release of cytokines such as platelet-derived growth factor (PDGF), transforming growth factor β1 (TGFβ1) and vascular endothelial growth factor (VEGF) achieving peak level at 14th day coinciding with cell ingrowth. PRF enhances the proliferation of various cell types, stimulates cellular differentiation and supplements the angiogenesis. Lastly, the presence of leucocytes and cytokines along with small amounts of lymphocytes in PRF can play a significant role in the self-regulation of inflammatory and infectious phenomenon [45].

In the present case, freshly prepared PRF fragments were placed inside the canal space without inducing bleeding as a part of the planned revascularization procedure. There is a lack of evidence regarding handling and placement of PRF in the root canal space. From clinical experience, placement of PRF fragments incrementally is more convenient than placing the membrane as a whole [20]. Another benefit of using a different scaffold than the blood clot would be the placement of MTA that could be more idealized. MM-MTA (Micro-Mega, Besançon, France) that has been shown to have excellent sealing properties [46] was placed directly over PRF to obtain a coronal seal.

Most of the case reports/series have shown an increase in root length and dentinal wall thickening with a reduction in the volume of the canal space radiographically [1, 2, 14 - 21]. Clinically the teeth were symptom free. This case report showed a similar clinic and radiographic outcome. However, where histological analysis of teeth, which had undergone revascularization treatment, was possible, the mineralized layer on the walls, which was observed, appeared to be of periodontal origin rather than pulpal origin. This histological outcome was observed in two case reports one of which had been revascularized by simple blood clot [47] and the other had used platelet-rich plasma [48]. This suggests that the new tissue was not of dentinogenic origin and highlights the shortcomings of dental radiography for characterizing new mineralized tissue in endodontics [49].

From a clinical perspective, the true origin of this tissue may not matter [49]. If the objective is to induce healing of the periapical tissues, stimulate bone regeneration and render the patient free from signs and symptoms, then this current case report can be deemed a clinical success. However, it may not exhibit “true” biological pulp regeneration but the filling of the root canal space with a vital biological tissue, which has further repair and defense capabilities. This represents a step forward when compared to gutta-percha or MTA [49]. Whether this revascularization process improves the root strength remains to be seen.

**Conclusion**

Platelet-rich fibrin is a potential scaffold in pulp revascularization procedures, as it is rich in growth factors, enhances cellular proliferation and differentiation, augments angiogenesis, acts as a matrix for tissue ingrowth, regulates inflammation reactions and has anti-infective properties. Additionally, it acts as an excellent matrix to support MTA placement. The disadvantages of using PRF include the requirement of special equipment, the invasive procedure of drawing blood in young patients, difficulty in handling and placing inside the canal space and increased cost of treatment. However, long-term clinical trials and histological studies are required to analyze the benefits of using PRF in revascularization/regeneration procedures.
References


